

The Use of the First Order System Transfer Function in the Analysis of Proboscis Extension Learning of Honey Bees, *Apis mellifera* L., Exposed to Pesticides

Charles I. Abramson · Igor I. Stepanov

Received: 10 August 2011 / Accepted: 28 December 2011 / Published online: 5 January 2012
© Springer Science+Business Media, LLC 2012

Abstract No attempts have been made to apply a mathematical model to the learning curve in honey bees exposed to pesticides. We applied a standard transfer function in the form $Y = B3 \cdot \exp(-B2 \cdot (X - 1)) + B4 \cdot (1 - \exp(-B2 \cdot (X - 1)))$, where X is the trial number; Y is proportion of correct responses, $B2$ is the learning rate, $B3$ is readiness to learn and $B4$ is ability to learn. Reanalyzing previously published data on the effect of insect growth regulators tebufenozide and diflubenzuron on the classical conditioning of proboscis extension, the model revealed additional effects not detected with standard statistical tests of significance.

Keywords Mathematical model · Pesticides · Honey bees · Learning curve

This paper describes a mathematical model of the learning process suitable for studies of proboscis conditioning (PER) in honey bees when bees are exposed to agro-chemicals such as pesticides. Although procedural variations exist in the way laboratories use the PER paradigm (Abramson et al. 2011), proboscis conditioning is widely used to investigate the

influence of pesticides and repellents on honey bee learning (Decourtye and Pham-Delegue 2002). In the PER paradigm, harnessed bees receive odor (the conditioned stimulus) and sucrose (the unconditioned stimulus) pairings. After several pairings, the proboscis extends to the conditioned stimulus (CS) prior to the presentation of the unconditioned stimulus (US). Honey bees have been exposed to pesticides (or repellents) by imbedding them in the conditioned stimulus (Abramson et al. 2006), serving as either the conditioned stimulus or unconditioned stimulus (Abramson et al. 2010), and pre-exposing the honey bee with a pesticide prior to conditioning (Abramson et al. 2004). Surprisingly, no attempts to mathematically characterize the learning process following exposure to agrochemicals have been previously reported.

Mathematical models have much to recommend for the evaluation of pesticides and repellents. Mathematical models of the learning process conveniently summarize large amounts of data, guide research and theory, and can characterize the results of various agro-chemical treatments across laboratories, species, and conditions. Mathematical models provide important information above and beyond that provided by standard significance testing.

Materials and Methods

Over the past several years, we have been developing a model of the learning process based on the first order system transfer function in the form $Y = B3 e^{-B2(X-1)} + B4 (1 - e^{-B2(X-1)})$ where X is the trial number and Y is the proportion of correct responses. The model contains three parameters: $B2$ —the learning rate; $B4$ —the asymptotic value of correct responses at $X = \text{Infinity}$ and

C. I. Abramson (✉)
Department of Psychology, Oklahoma State University,
116 N. Murray, Stillwater, OK 74078, USA
e-mail: charles.abramson@okstate.edu

I. I. Stepanov
Department of Neuropharmacology, Institute for Experimental
Medicine, 12 Acad. Pavlov Street, St. Petersburg 197376, Russia

B3—the value of correct responses at the beginning of training (i.e. $B3 = Y$ at $X = 1$). Since $B2 = 1/\tau$, it means that $1/B2$ is the number of trials required for achievement of 63% from the difference between B3 and B4.

We treat B3 as an estimator of the functional state of an animal at the beginning of training that includes, in particular, the value of prior learning based on previous experience and readiness to learn. Since the independent variable is the value of correct responses (i.e. a proboscis extension to the CS) the greater the value of B3, the higher is the readiness to learn. Thus, B3 is the ‘readiness to learn’.

The learning mechanisms are reflected in the learning rate (B2) and asymptotic level (B4). In the case of PER conditioning, B4 is the maximal possible number of correct responses after a very large number of trials. The greater the value of B4, the higher is the ability to learn. Our rationale for using coefficient B4 is that the number of correct responses does not always increase to 1 (or 100%). That is why, in our view, it is impossible to pre-fix B4 to 1.0. Below we will designate B4 as ‘the ability to learn’.

Additionally, we restricted the value of coefficient B2. Coefficient B2 must exceed zero by definition of the mathematical model. The upper limit we established for B2 is equal to 5.0. It is known that the first order system reaches 99.3% of its asymptotic value during five time constants (Grodins 1963; Milsum 1966). Since $B2 = 1/\tau$, it means that if $B2 = 5$, then an animal achieves its asymptotic maximum of conditioned responses in one session. In other words, we suppose that the exponential learning curve exists, if $0.001 < B2 < 5.0$.

To illustrate the type of information obtained from our model we re-analyzed some of our previously published data on the effect of insect growth regulators Confirm[®]2F (Tebufenozide) and Dimilin[®] (Diflubenzuron) on the acquisition of honey bee PER learning (Abramson et al. 2004). In that study we concluded that exposure to Confirm[®]2F or Dimilin[®] affected the acquisition of learning. Honey bees were fed using a microsyringe 10 μ l of Confirm[®]2F or Dimilin[®] 10 min prior to conditioning. Those in the Confirm[®]2F group received either 0, 16 μ g/bee, 24 μ g/bee, 32 μ g/bee, 69.4 μ g/bee, or 131 μ g/bee of Tebufenozide. Honey bees in the Dimilin[®] group received either 0 μ g/bee, 3.4 μ g/bee, 8.5 μ g/bee, 16 μ g/bee, 32 μ g/bee, or 69.4 μ g/bee of Diflubenzuron. Each group contained 26 bees.

The conditioned stimulus was a 3 s presentation of cinnamon odor and the unconditioned stimulus a 2 s feeding of 1.8 M sucrose. Each animal received 12 acquisition trials. The time between trials was 10 min. The CS was presented first, and when the CS terminated the US was presented. Using ANOVA procedures we previously showed that both Confirm[®]2F and Dimilin[®] influenced the acquisition of honey bee learning. However, when we

apply the model to this data set much more information can be extracted.

Results and Discussion

Effect of Confirm[®]2F on Acquisition of the PER

Exposure to Confirm[®]2F significantly decreased the learning rate (B2) in each concentration especially in bees given 32 μ g/bee. Exposure to 69.4 μ g/bee increased readiness to learn (B3) but this was not the case at any other concentration. Ability to learn (B4) was reduced in animals given concentrations of 24 and 131 μ g/bee but not at other concentrations.

Effect of Dimilin[®] on Acquisition of the PER

With the exception of bees exposed to 16 μ g/bee, exposure to Dimilin[®] significantly decreased the learning rate (B2). The decrease was significantly pronounced in bees exposed to 32 μ g/bee. Dimilin[®] also significantly decreased the ability to learn (B4) at each concentration, especially in bees receiving 24 μ g/bee and 69.4 μ g/bee. Dimilin[®] did not influence on readiness to learn (B3) at any concentration.

Comparison of Effects of Both Pesticides

The main effect of Confirm[®]2F was to decrease the learning rate (B2). In contrast, the main effect of Dimilin[®] was to decrease the ability to learn (B4). These results suggest that Dimilin[®] is more dangerous to honey bees than Confirm[®]2F even though both insect growth regulators are considered “harmless” to honey bees. Readiness to learn (B3) was not affected in honey bees with the exception that bees given 69.4 μ g/bee of Confirm[®]2F increased their readiness to learn.

It should be noted that fitting the learning curve with the first order system transfer function is not the only statistically sound approach to analyzing learning data. Any statistical technique that both adjusts for the inherent correlation structure and accounts for both the dichotomous data and the pattern of learning over time is appropriate. This includes “growth curve” analyses with asymptotic, increasing or decreasing functions.

Hartz et al. (2001) proposed a model based on logistic growth curve analysis for PER learning in honey bees. One rationale for the use of the logistic growth curve model is that it is available in many statistical packages and no sophisticated programming is necessary. A comparison of the logistic growth curve model and the first order transfer model advocated here revealed that our model provided better fits for their data and that our model can extract more

information from the data in the form of our three coefficients. Our model also has the advantage that it has been successfully applied to learning in many animals including land snails (Balaban and Stepanov 1996), rats (Stepanov and Abramson 2008) and humans (Stepanov and Abramson 2005) including Luriya's test of learning Russian words (Abramson and Stepanov 2007).

Of particular interest is that we have used the model to characterize patients suffering from multiple sclerosis (Stegen et al. 2010), for patients with type 2 Diabetes Mellitus (Stepanov et al. 2010) and to characterize the learning of children given the California Verbal Learning Test—(Stepanov et al. 2011). We believe that this is an advantage of our model because it can be used to directly compare performance decrements produced by a variety of environmental factors and disease including environmental contaminants and to compare coefficients across species.

As with any model, extensive validation is needed. As discussed above our model has been validated with a number of organisms in a variety of situations. This is the first paper, however, that attempts to apply the model to pesticide data. It should be noted that to apply the model, a minimum of 7 conditioning trials must be used. Several studies in the pesticide literature use only 3 conditioning trials (El Hassani et al. 2005; Han et al. 2010) and so few trials do not supply the model with enough information for accurate modeling.

We believe that our model can be applied to many aspects of honey bee learning as it relates to pesticide exposure. Of particular interest is an examination of extinction (when the CS is no longer followed by the US) and discrimination learning (when two CSs are used with one of them paired with a US). An agro-chemical, for example, may not influence the acquisition of learning but its persistence when the US or reward is discontinued. Agro-chemicals may also affect the ability of honey bees to discriminate between stimuli.

In addition to classical conditioning, the model has been used to characterize the maze performance of rats exposed to various pharmacological agents (Stepanov and Abramson 2008) and we hope to expand the model to other areas of learning such as operant conditioning (Abramson 1994). We have not yet extended the first order system transfer function to include other dependent variables such as flight time.

Of particular interest is the application of the model to characterize so-called “green products” such as essential oil based pesticides which often make claims of being harmless to honey bees (Abramson et al. 2006). Our model opens the possibility of characterizing these, and other agro-chemicals, by their effect on learning rate (B2), readiness to learn (B3) and ability to learn (B4). These

parameters provide additional information on the effect of an agro-chemical that standard significance tests cannot provide. For example, the application of the model to the insect growth regulators Dimilin® and Confirm® 2F reveal that Dimilin® is more dangerous to honey bees because it affected the bees' ability to learn (parameter B4) although both growth regulators are considered “harmless.”

Details on how to use the model including programming language for SPSS is available in Stepanov et al. (2010) and directly from the authors. We would be glad to assist any researcher who wants to apply the model to their data.

References

- Abramson CI (1994) A primer of invertebrate learning: the behavioral perspective. American Psychological Association, Washington DC
- Abramson CI, Stepanov II (2007) The application of a mathematical model for the assessment of the memorization curve in drug addicts and alcoholics. *J Drug Addict Educ Erad* 3:239–262
- Abramson CI, Squire J, Sheridan A, Mulder PG Jr (2004) The effect of insecticides considered harmless to honey bees (*Apis mellifera* L.): proboscis conditioning studies using the insect growth regulators confirm® 2F (tebufenozide) and dimilin® (diflubenzuron). *Environ Entomol* 33:378–388
- Abramson CI, Singleton JB, Wilson MK, Wanderley PA, Ramalho FS, Michaluk LM (2006) The effect of an organic pesticide on mortality and learning in Africanized honey bees (*Apis mellifera* L.) in Brasil. *Am J Environ Sci* 2:37–44
- Abramson CI, Giray T, Mixson TA, Nolf SL, Wells H, Kence A, Kence M (2010) Proboscis conditioning experiments with honey bees (*Apis mellifera caucasica*) show butyric acid and deet not to be repellents. *J Insect Sci* 10:122 available online: insect-science.org/10.122
- Abramson CI, Sokolowski MBC, Wells H (2011) Issues in the study of proboscis conditioning. In: Columbus F (ed) Social insects: structure, function, and behavior. Nova Science Publishers, Hauppaug, pp 25–49
- Balaban PM, Stepanov II (1996) Innate and acquired behavior of mollusks. In: Abramson CI, Shuranova ZP, Burmistrov YM (eds) Russian contributions to invertebrate behavior. Praeger Publishers, Westport, pp 77–109
- Decourtaye A, Pham-Delegue MH (2002) The proboscis extension response: assessing the sublethal effect of pesticides on the honey bee. In: Devillers J, Pham-Delegue MH (eds) Honey bees: estimating the environmental impact of chemicals. Taylor & Frances, London, pp 67–84
- El Hassani AK, Dacher M, Gauthier M, Armengaud C (2005) Effects of sublethal doses of fenopril on the behavior of the honeybee (*Apis mellifera* L.). *Pharmacol Biochem Behav* 82:30–39
- Grodins FS (1963) Control theory and biological systems. Columbia University Press, New York
- Han P, Niu C-Y, Lei C-L, Cui J-J, Desneux N (2010) Use of an innovative T-tube maze assay and the proboscis extension response assay to assess sublethal effects of GM products and pesticides on learning capacity on the honey bee *Apis mellifera* L. *Ecotoxicology* 19:1612–1619
- Hartz SM, Ben-Shahar Y, Tyler M (2001) Logistic growth curve analysis in associative learning data. *Anim Cogn* 4:185–189
- Milsum JH (1966) Biological control systems analysis. McGraw-Hill, New York

- Stegen S, Stepanov I, Cookfair D, Schwartz E, Hojnacki D, Weinstock-Guttman B, Benedict RHB (2010) Validity of the California verbal learning test-II in multiple sclerosis. *Clin Neuropsychol* 24:189–202
- Stepanov II, Abramson CI (2005) A new mathematical model for assessment of memorization dynamics. *Span J Psychol* 8:142–156
- Stepanov II, Abramson CI (2008) The application of the first order system transfer function for fitting the 3-arm radial maze learning curve. *J Math Psychol* 52:309–319
- Stepanov II, Abramson CI, Wolf OT, Convit A (2010) The application of the first order system transfer function for fitting the California verbal learning test learning curve. *JINS* 16:443–452
- Stepanov II, Abramson CI, Warschausky S (2011) Assessment of the learning curve from the California verbal learning test—children's version with the first-order system transfer function. *Child Neuropsychol* 17:330–346